



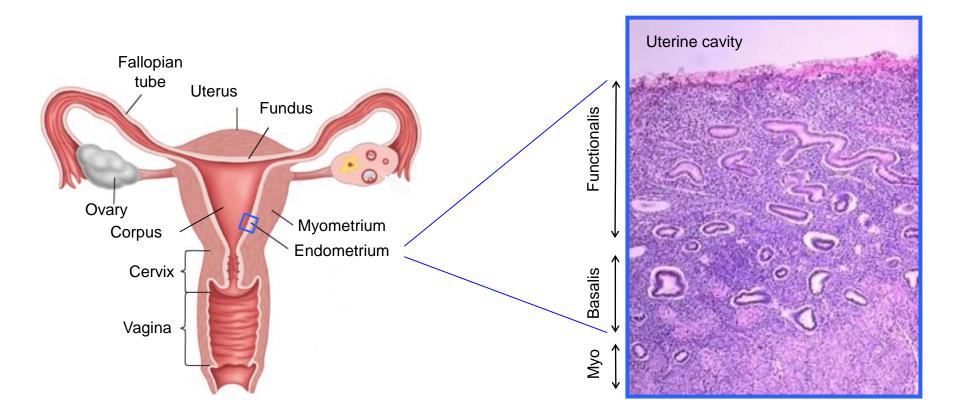
Endocrine and paracrine control of menstruation

Patrick Henriet Institut de Duve, UCLouvain

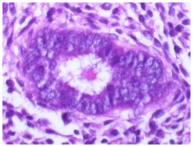


CHU Saint Pierre November 19, 2022

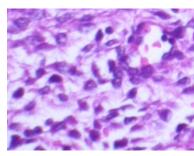
Anatomy and histology of the human endometrium





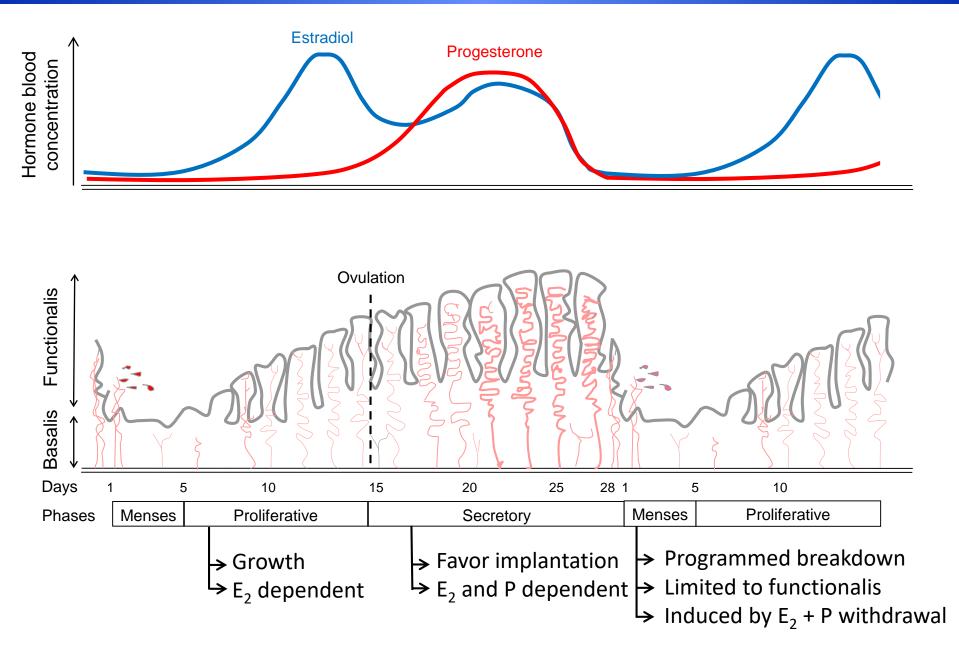


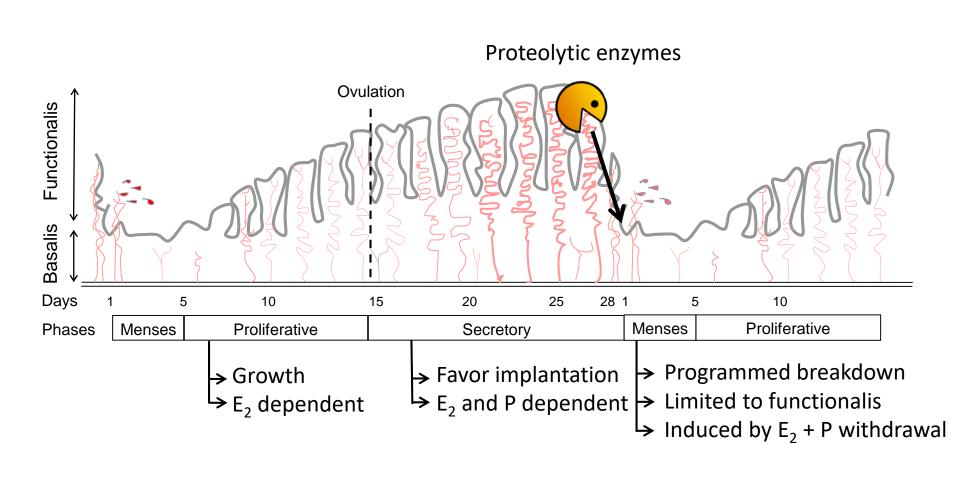
Stroma



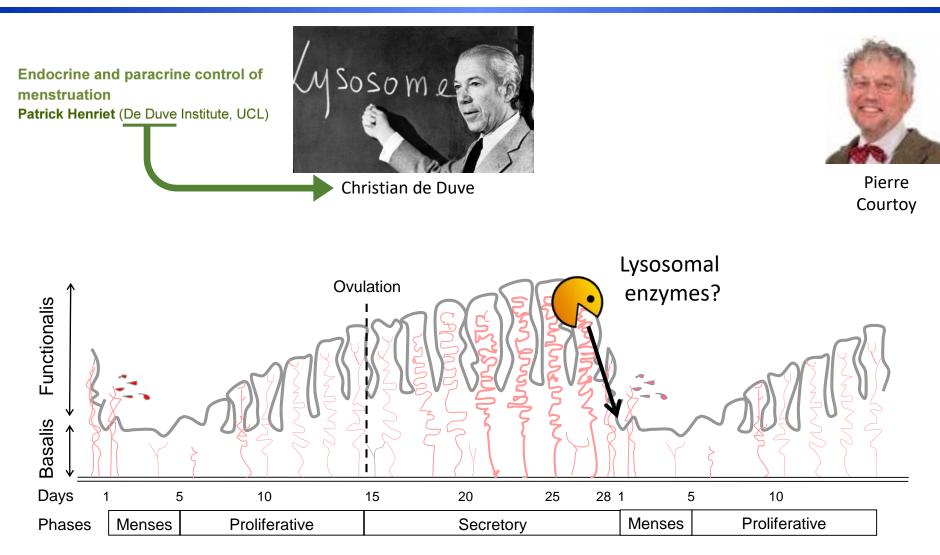
- Fibroblasts, vessels, inflammatory cells
- Extracellular matrix, rich in collagens

The menstrual cycle

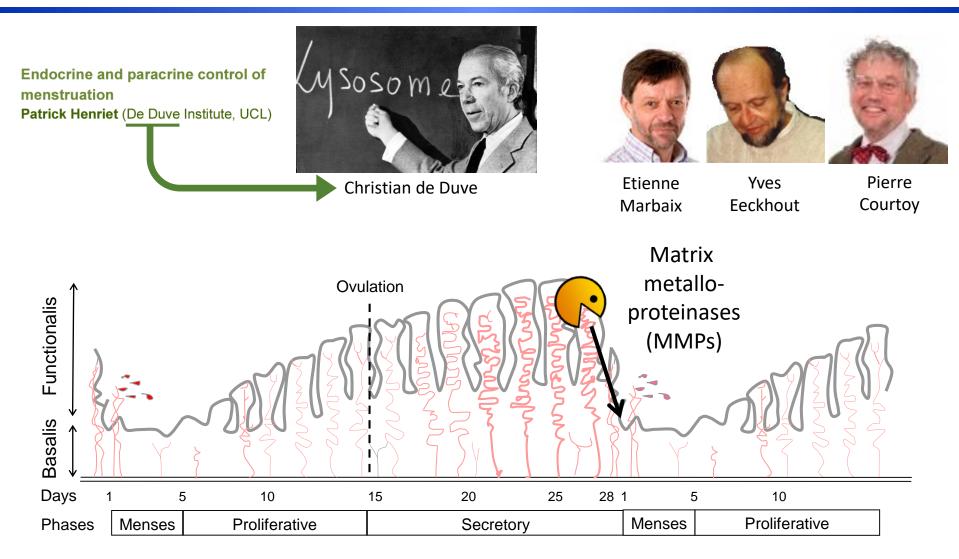




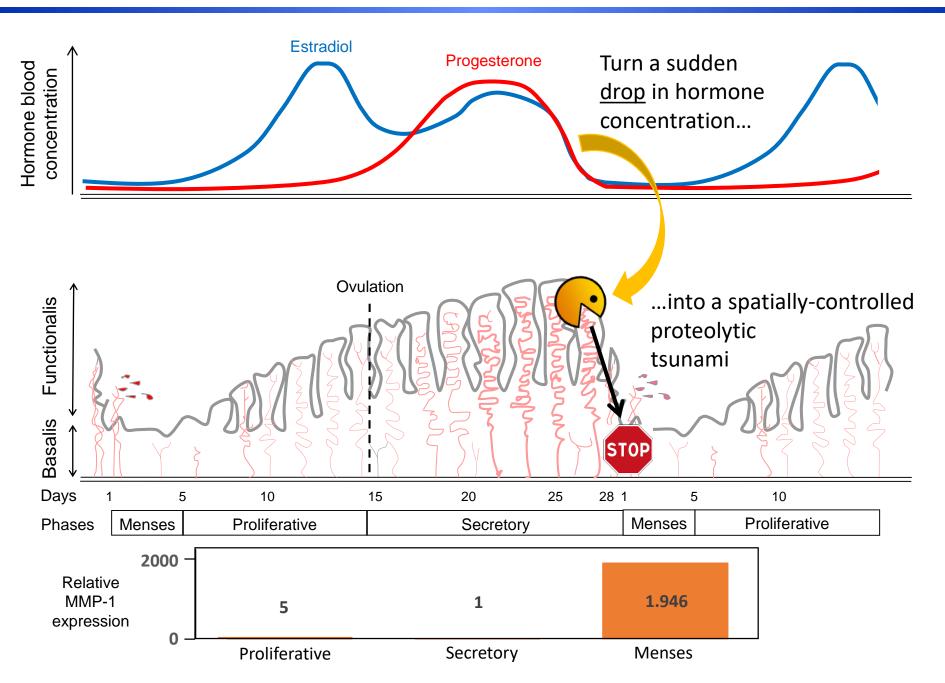
Enzymes responsible for tissue breakdown during menstruation?

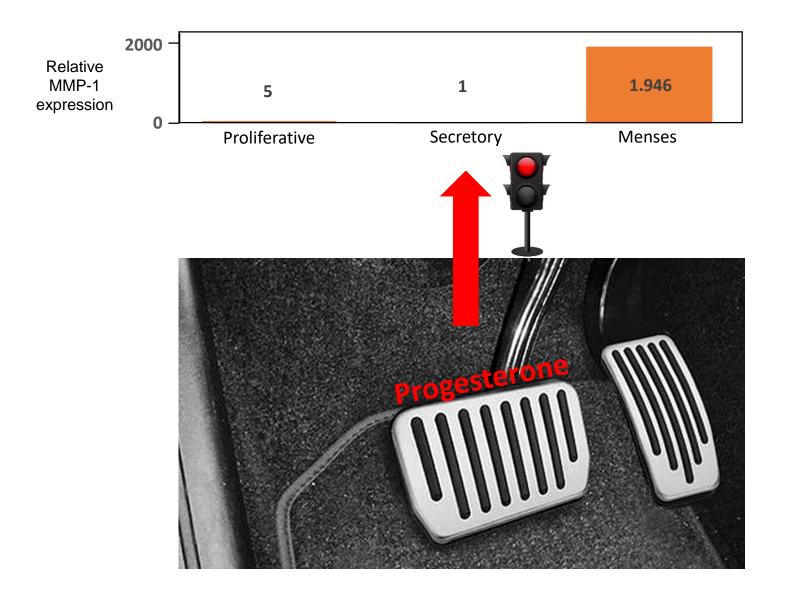


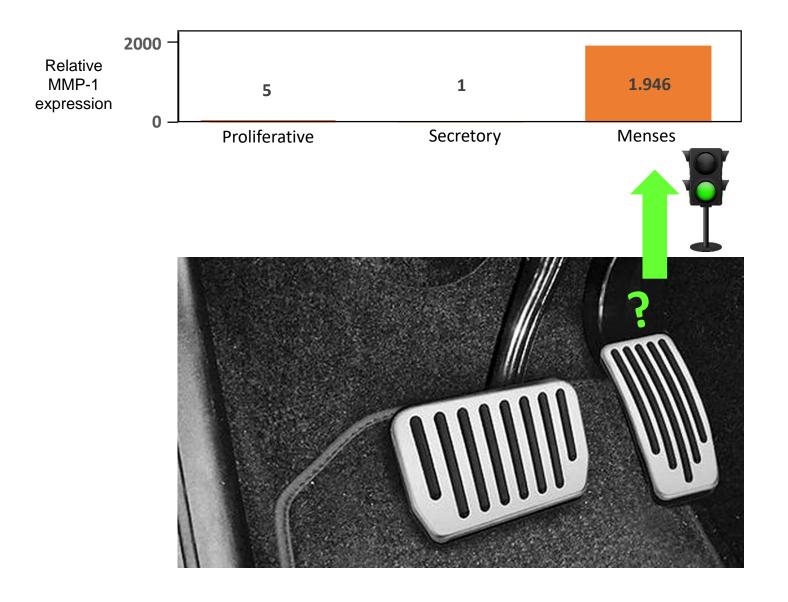
Enzymes responsible for tissue breakdown during menstruation?



The central question in understanding regulation of menstruation







- Menstruating species :
 - > only humans and closely-related primates, with few remarkable exceptions



elephant shrew

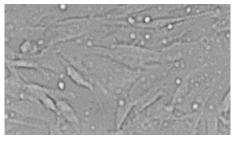


some bats

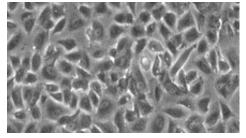


a rodent : Acomys

- spontaneous ovulation followed by decidualization during the secretory phase
- Decidualization : animal models => a prerequisite for human menstruation



elongated fibroblasts



=> round secretory decidual cells

- induced by the production of cAMP in combination with progesterone
- characterized by production of prolactin, glycogen, IGFBP1 and FOXO-1A

BUT HOW DECIDUALIZATION INFLUENCES MENSTRUATION REMAINS UNCLEAR

FLSEVIEF

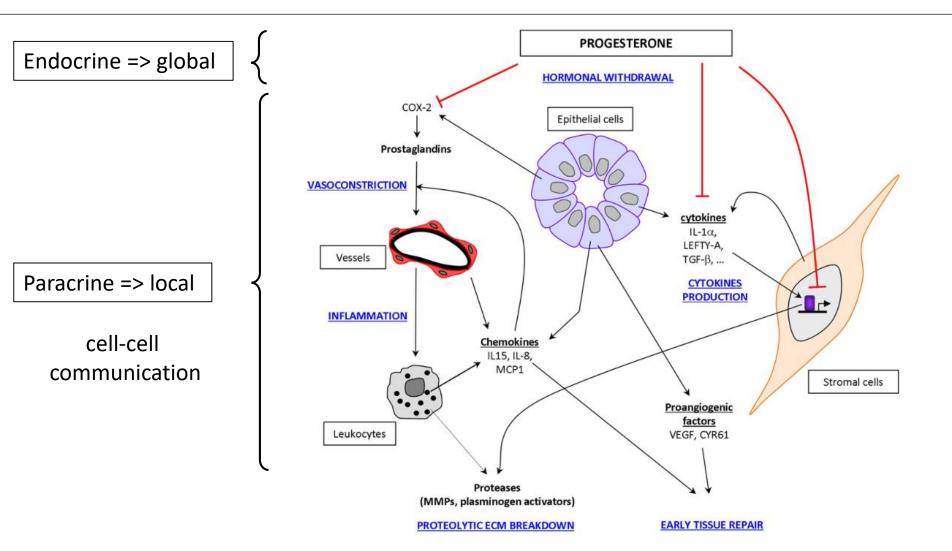
Review

The endocrine and paracrine control of menstruation Patrick Henriet^{*,1}, Héloïse P. Gaide Chevronnay¹, Etienne Marbaix

Cell Biology Unit, de Duve Institute, Université catholique de Louvain, avenue Hippocrate, 75, B-1200 Bruxelles, Belgium

Contents lists available at SciVerse ScienceDirect Molecular and Cellular Endocrinology

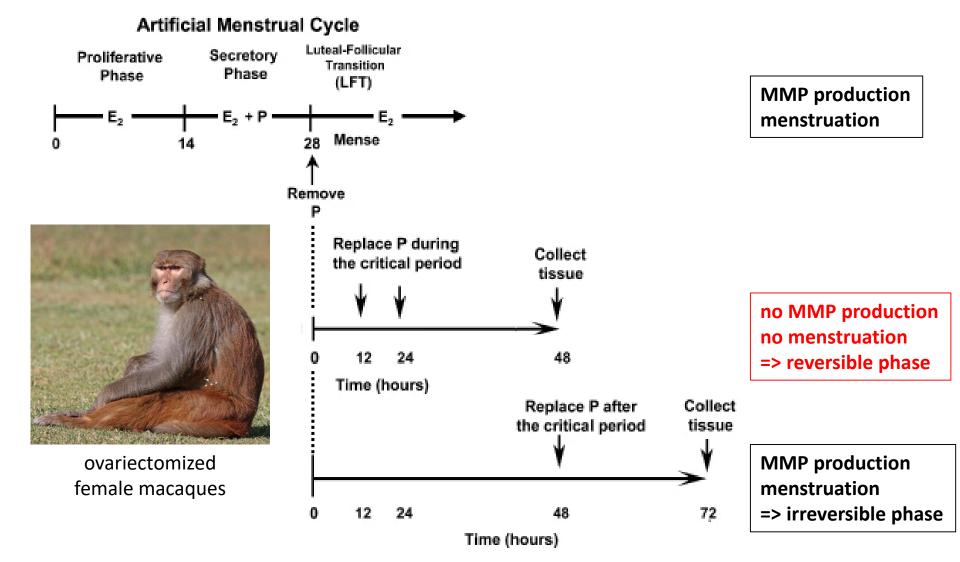
journal homepage: www.elsevier.com/locate/mce



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1) Progesterone withdrawal is sufficient

=> lessons from the non-human primate model (Slayden et al., 2006)

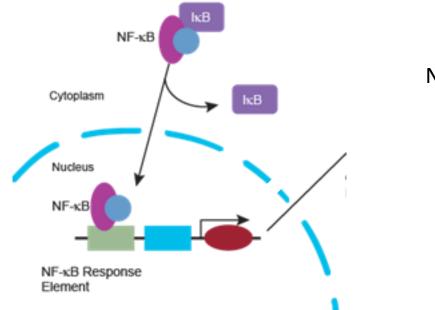


Reversible phase (if P back <24h in non-human primate model)

Irreversible phase (when P back >36h in non-human primate model)

Reversible phase (if P back <24h in non-human primate model)

2) Increased production of ROS => increased production and activation of $NF-\kappa B$



NF-κB (nuclear factor kappa B)

transcription factor

key role in inflammation

Menstruation is a physiological model of self-limiting inflammation

Hilary Critchley, 2020

Reversible phase (if P back <24h in non-human primate model)

2) Increased production of ROS => increased production and activation of <u>NF-κB</u>

3) Increased local expression of :

- inflammatory mediators such as COX-2 => increased prostaglandin production
- cytokines such as IL-1, IL-6, TNF => induction of MMP expression
- chemokines such as IL-8, CCL-2/MCP-1 => monocyte/macrophage recruitment

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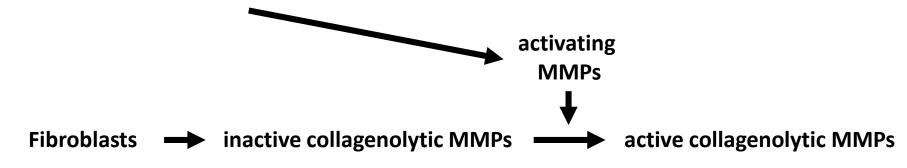
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5) Vasoconstriction and hypoxia

Contribution to menstruation seems modest

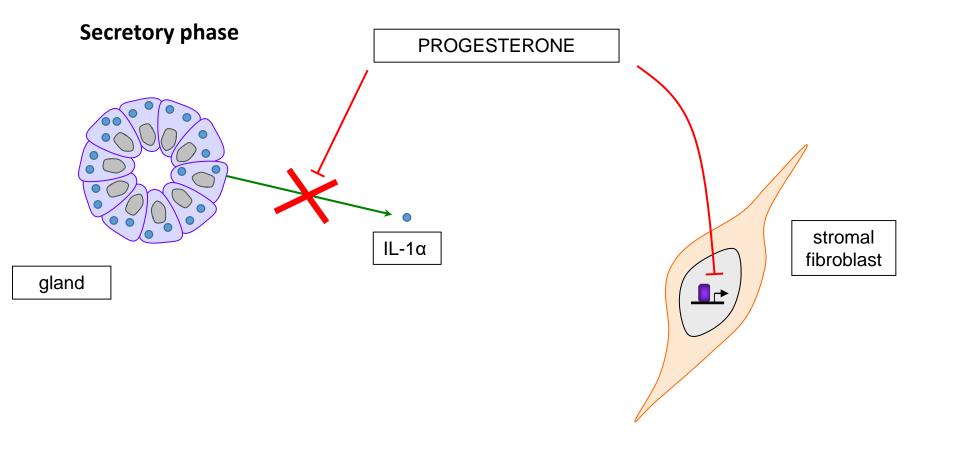
Prevents excessive blood loss during menstruation

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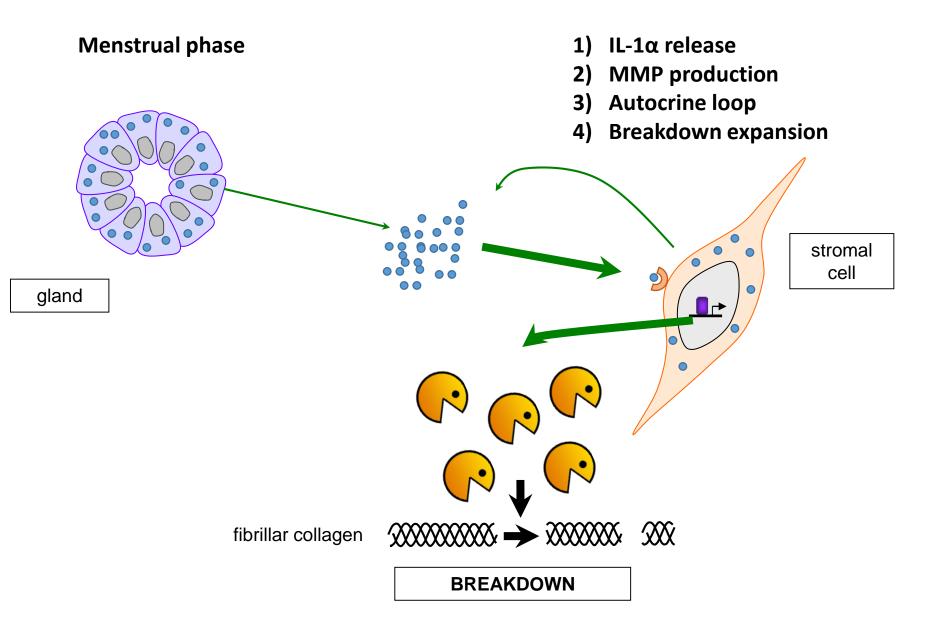
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- 4) Cellular influx : neutrophils : 10% (proliferative) → 20-45% stromal cells (premenstrual)
- 5) Vasoconstriction and hypoxia
- 6) Vessel permeability and fragility
- 7) Focal expression and activation of MMPs => tissue breakdown





Cornet, 2002, 2005 ; Gaide Chevronnay, 2008 ; Singer, 1997 ; Pretto, 2008



Cornet, 2002, 2005 ; Gaide Chevronnay, 2008 ; Singer, 1997 ; Pretto, 2008

• Non-human primate model

- MMP production stops after a few days
- estradiol is not required
- Macrophages endocyte and clear cellular debris
- Repair and regeneration : precise contribution of stem cells?
- **Hypoxia** => hypoxia-inducible factor (HIF)

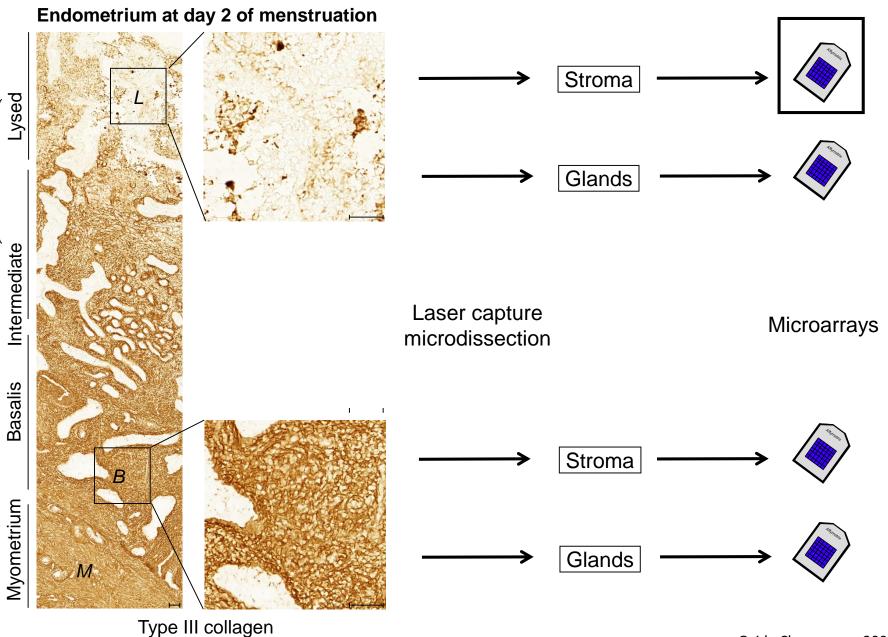
=> vascular endothelial growth factor (VEGF)

=> neovascularisation

- Fragments from the functionalis
 - express genes involved in tissue regeneration <u>during</u> menstruation
 - could contribute to regenerating a new functionalis if not cleared
 - > and to pathogenesis of **endometriotic lesions** through retrograde menstruation

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Gaide Chevronnay, 2009



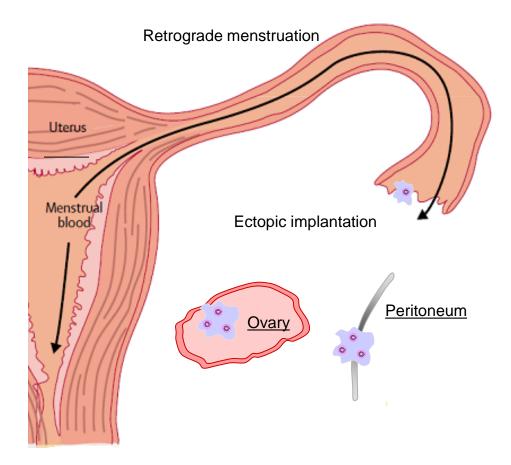
Functionalis

EXTRACELLULAR MATRIX DEGRADATION

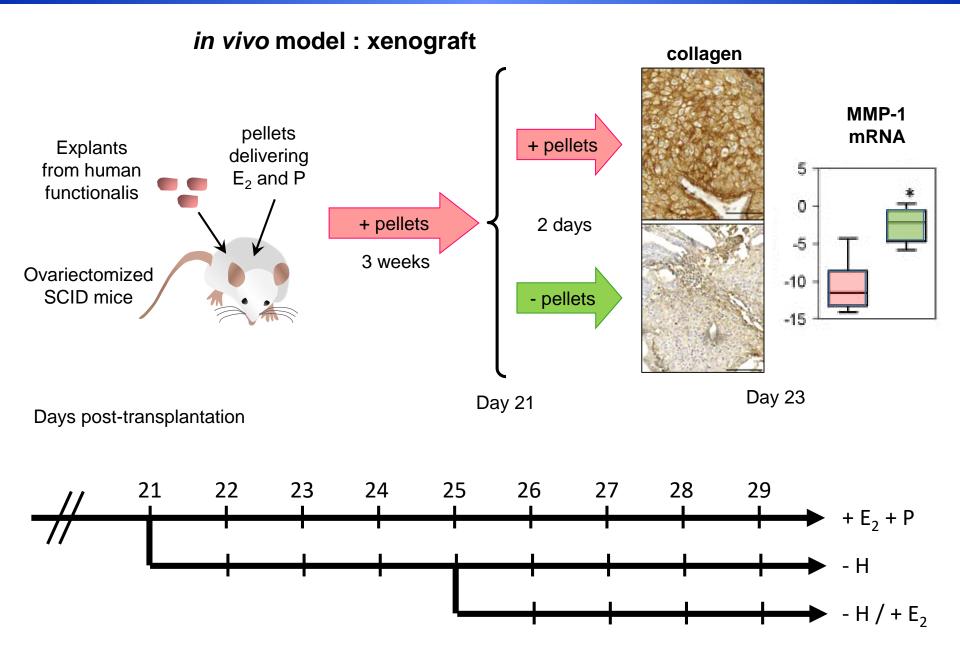
Symbol	Description
ANGPT2	angiopoietin 2
BGN	biglycan
CHI3L1	chitinase 3-like 1 (cartilage glycoprotein-39)
CILP	cartilage intermediate layer protein, nucleotide pyrophosphohydrolase
COL12A1	collagen, type XII, alpha 1
COL18A1	collagen, type XVIII, alpha 1
COL1A1	collagen, type I, alpha 1
COL24A1	collagen, type XXIV, alpha 1
COL5A2	collagen, type V, alpha 2
COL7A1	collagen, type VII, alpha 1
CTHRC1	collagen triple helix repeat containing 1
ENTPD4 / LOXL2	ectonucleoside triphosphate diphosphohydrolase 4 / lysyl oxidase-like 2
EPYC	epiphycan
GNLY	granulysin
GPC1	glypican 1
GZMB	granzyme B (granzyme 2, cytotoxic T-lymphocyte-associated serine esterase 1)
IGFBP1	insulin-like growth factor binding protein 1
IGFBP6	insulin-like growth factor binding protein 6
IL8	interleukin 8
INHBA	inhibin, beta A
ITGA2	integrin, alpha 2 (CD49B, alpha 2 subunit of VLA-2 receptor)
LAMC3	laminin, gamma 3
LOX	lysyl oxidase
LTBP2	latent transforming growth factor beta binding protein 2
MMP1	matrix metallopeptidase 1 (interstitial collagenase)
MMP10	matrix metallopeptidase 10 (stromelysin 2)
MMP12	matrix metallopeptidase 12 (macrophage elastase)
MMP3	matrix metallopeptidase 3 (stromelysin 1)
MMP9	matrix metallopeptidase 9 (gelatinase B)
NID2	nidogen 2 (osteonidogen)
P4HA2	prolyl 4-hydroxylase, alpha polypeptide II
P4HB	prolyl 4-hydroxylase, beta polypeptide
PCOLCE	procollagen C-endopeptidase enhancer
PLAT	plasminogen activator, tissue
PLAU	plasminogen activator, urokinase
PLAUR	plasminogen activator, urokinase receptor
PLOD2	procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2
PTHLH	parathyroid hormone-like hormone
SERPINA1	serpin peptidase inhibitor, clade A (alpha-1 antiproteinase, antitrypsin), member 1
STC1	stanniocalcin 1
SULF1	sulfatase 1
TFPI2	tissue factor pathway inhibitor 2
TIMP1	TIMP metallopeptidase inhibitor 1
TIMP3	TIMP metallopeptidase inhibitor 3
TNFAIP2	tumor necrosis factor, alpha-induced protein 2
TNFAIP6	tumor necrosis factor, alpha-induced protein 6
TNFRSF11B	tumor necrosis factor receptor superfamily, member 11b (osteoprotegerin)

EXTRACELLULAR MATRIX PROTECTION OR SYNTHESIS

Gaide Chevronnay, 2009

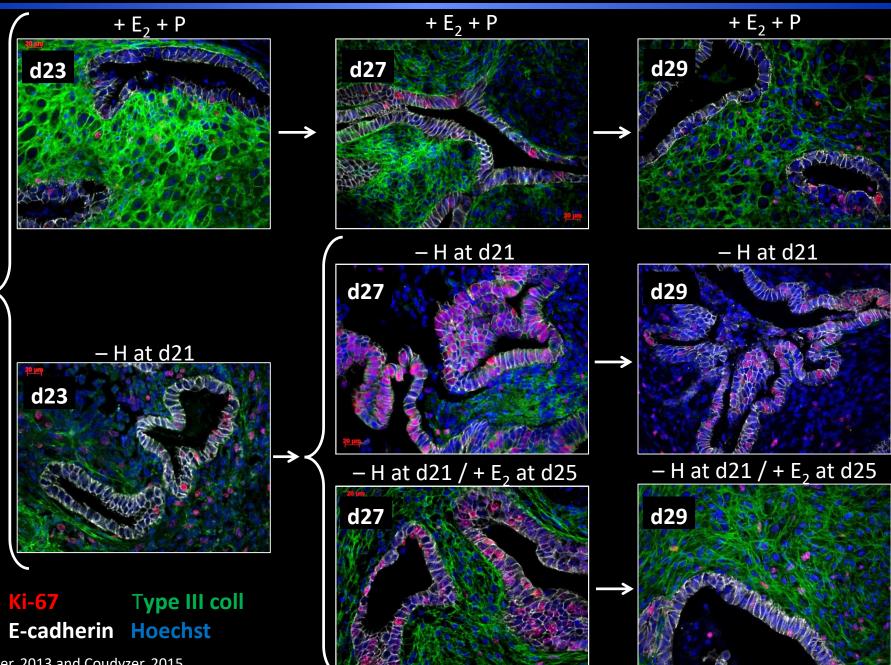


Regenerative potential of tissue fragments from the functionalis



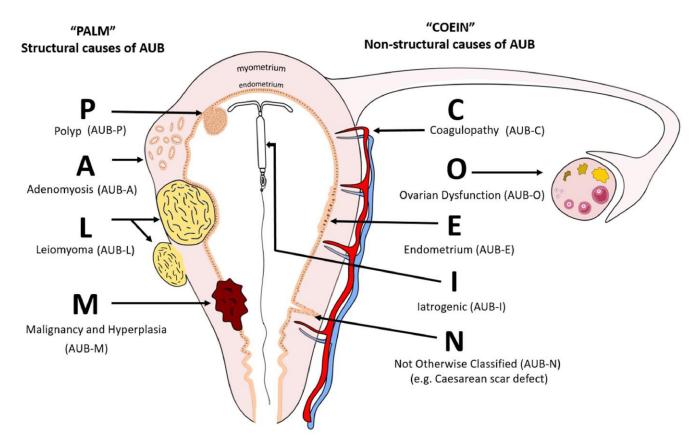
Coudyzer, 2013 and Coudyzer 2015

Hormone withdrawal is sufficient to induce cell proliferation



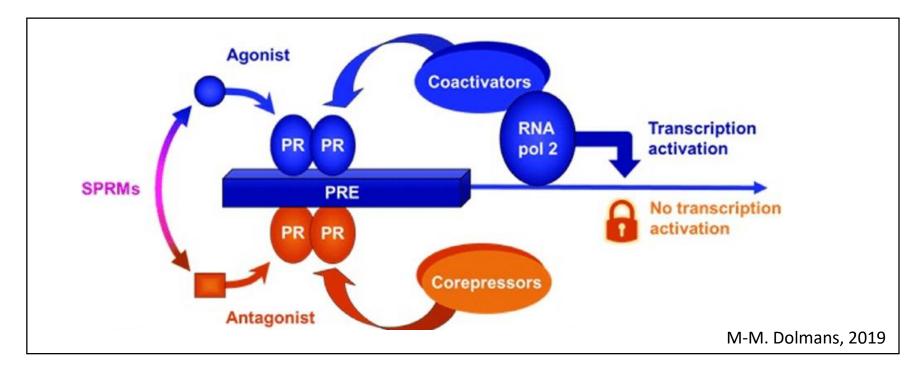
Coudyzer, 2013 and Coudyzer, 2015

• Multiple causes for AUB : the P A L M – C O E I N classification system



- Recurrent symptoms related with the sequence of events for menstruation
 - > untimely/excessive release of proinflammatory cytokines (IL-1 α)
 - inappropriate prostaglandin production => reduced vasoconstriction
 - untimely/excessive release of MMPs

- Favorite target for therapeutic approaches : progesterone receptor (PR)
 - selective progesterone receptor modulators (SPRM)
 - wide range of actions from perfect agonist to full antagonist
 - examples : mifepristone (RU486) ; ulipristal acetate



- <u>but</u> : potential long term side effects
- <u>but</u> : alternative pathways for progesterone response, PR-independent

Conclusions

• Chronological sequence of events preceding menstruation

- But correlation is not causation
 - mechanistic questions

- More research needed :
 - > to better understand the contribution of decidualization
 - > to improve identification of targets / drugs for AUB therapies



